lealt A Family of Medical Product

INTENDED USE

HealthLink Inc. Transport Medium (UTM[®]) System is intended for collection and transport to the analy sis laboratory of clinical specimens with suspected presence of viruses, chlamydiae, mycoplasmas o ureaplasmas for subsequent culture techniques.

SUMMARY AND PRINCIPLES

SUMMARY AND PRINCIPLES One of the routine procedures in the diagnosis of infections caused by viruses, chlamydiae, mycopla-smas or ureaplasmas involves collection and refrigerated transport of biological specimens. Using the UTM® System, the collected specimen can be stored for up to 48 hours at 2-25°C. The UTM® consists of a Hanks' Balanced Salt Solution (HBSS) enriched with proteins and sugars with a neutral pH and pH indicator. The medium contains some antibiotics and antimycotics to inhibit overgrowth of bacteria and yeasts, maintain cellular integrity and encourage preservation of viruses and chlamydiae if specimens are frozen at -70°C or colder until the time of processing^{2,3,6,13}.

PRODUCT DESCRIPTION

UTM® System is ready for use and requires no further preparation. It is available in the various confi-gurations listed in Table 1 and supplied in a labelled screw-cap test tube filled with different volumes of UTM®. The packaging in kits also includes a sterile collection device.

Catalog No.	Description	Pack Size
302C.HL	3 ml of UTM [®] medium in 16x100 mm screw-cap tube with internal shaped conical bottom Two regular size applicator swabs polyester tipped and molded breaking point at 100 mm	50 kits per package 6 x 50 kits per box
3C011N.HL	1 ml of UTM [®] medium in 12x80 mm screw-cap tube with internal shaped conical bottom. One regular size applicator swab with molded breaking point at 80mm	50 kits per package 6 x 50 kits per box
330C.HL	3 ml of UTM [®] medium in 16x100 mm screw-cap tube with internal shaped conical bottom.	50 tubes per package 6 x 50 tubes per box
3C030N.HL	1 ml of UTM [®] medium in 12x80 mm screw-cap tube with internal shaped conical bottom.	50 kits per package 6 x 50 kits per box
3C036N.HL	CO36N.HL 3 ml of UTM [®] medium in 16x100 mm screw-cap tube with internal shaped conical bottom. One flexible applicator swab with flocked nylon fiber and molded breaking point at 100 mm.	
3C037N.HL	3 ml of UTM [®] medium in 16x100 mm screw-cap tube with internal shaped conical bottom. One minitip applicator swab with flocked nylon fiber and molded breaking point at 100 mm.	50 kits per package 6 x 50 kits per box

1

HPC170A Rev.01 Date 2019.09

WARNINGS AND PRECAUTIONS

- Single-use device for professional in vitro diagnostic use. Do not use beyond the expiry date. Do not immerse the collection device in the UTM[®] before sampling.
- 2 3
- 4.
- Specimens for the search of viruses, chlamydiae, mycoplasmas and ureaplasmas must be collected and handled using personal protective equipment against biological risk according to published manuals and guidelines^{1,4,6,7,9,17}. Repeated freezing and thawing of specimens may reduce the recovery of viable
- 5 organisms.
- 6.
- organisms. Identify the test tube containing the specimen. Do not use if the device shows visible signs of damage or contamination, if you observe medium leaking from the test tube or if the medium appears murky yellow The use of this product in combination with diagnostic kits or instruments must be validated by the user prior to use. 8.

INSTRUCTIONS FOR USE

Proper collection of the specimen from the patient (e.g. aspirates, small tissue or faecal specimens, urine) is a crucial aspect for successful isolation and identification of infectious organisms

In order to maintain optimal microorganism viability, transport the specimens to the labo-ratory as soon as possible considering that the viral concentrations reach the maximum values during the acute phase of the disease.

- UTM® in KIT
 Open the UTM® kit package and remove the medium test tube and the internal bag containing the sterile swab.
 Take the sterile swab out of its bag and collect the clinical specimen; to prevent the risk of contamination, make sure that the swab tip comes into contact with the unit of the optic.
 - 3
 - Collection site only. After collecting the specimen, unscrew and remove the cap from the test tube taking care not to spill the medium. Insert the swab into the test tube until the breakpoint is level with the test tube 4.
 - opening. Bend and break the swab at the breakpoint holding the test tube away from your 5.

 - being and bleak the swap at the breakpoint holding the test tube away norm your face and discard the excess part. Screw the cap back onto the test tube and hermetically seal it. Process the specimen contained in the UTM® within 48 hours from collection storing the test tube at $2-25^{\circ}$ C. 6
 - 8. Before processing, vortex for 20 seconds in order to encourage specimen release from the swab and homogenize the medium.

- UTM® in bulk

 1.
 After collecting the specimen, unscrew and remove the cap from the UTM® test tube taking care not to spill the medium.
 - Insert the previously validated swab into the test tube until the breakpoint (if present) is level with the test tube opening. Bend and break the swab at the breakpoint holding the test tube away from your face; should the swab used not have a breakpoint, cut the excess part of the shaft and di-coard it 3.
 - scard it
 - 4 5
 - scard it. Screw the cap back onto the test tube and hermetically seal it. Process the specimen contained in the UTM® within 48 hours from collection storing the test tube at 2-25°C. Before processing, vortex for 20 seconds in order to encourage specimen release from 6. the swab and homogenize the medium.

If processing is delayed (over 48 hours), the specimens must be frozen at -70°C or colder.

3C038N.HL	3 ml of UTM [®] medium in 16x100 mm screw-cap tube with internal shaped conical bottom. One regular applicator swab with flocked nylon fiber and molded breaking point at 100 mm.	50 kits per package 6 x 50 kits per box
3C039N.HL	 3 ml of UTM[®] medium in 16x100 mm screw-cap tube with internal shaped conical bottom. One flexible applicator swab with flocked nylon fiber and molded breaking point at 100 mm. One regular applicator swab with flocked nylon fiber and molded breaking point at 100 mm. 	50 kits per package 6 x 50 kits per box
3C040N.HL	1 ml of UTM [®] medium in 12x80 mm screw-cap tube with internal shaped conical bottom. One flexible applicator swab with flocked nylon fiber and molded breaking point at 100 mm.	50 kits per package 6 x 50 kits per box

Table 1: product description

REAGENTS The UTM® formulation includes proteins for virus stabilization¹⁷, antibiotics and antimycotics to prevent overgrowth of bacterial and fungal flora and a buffer solution to maintain a neutral pH

Components	Quantity g/liter		
Sugars	50-100 g/l		
HBSS solution	5-20 g/l		
Bovine serum albumin	5-20 g/l		
Buffered solution	5-20 g/l		
Jelly	1-5 g/l		
Amino acids	< 1 g/l		
Antibiotics	< 1 g/l		
PH indicator	< 1 g/l		
pH 7,3 ±	0,2 a 2÷25 °C		

REQUIRED MATERIALS BUT NOT PROVIDED Materials suitable for isolation, differentiation and culture of viruses, chlamydiae, mycoplasmas and ureaplasmas. The collection device is not provided in packaging in bulk.

STORAGE The product must be stored in its original packaging at a temperature between 2 and 25°C until the time of use. Do not overheat or freeze prior to use

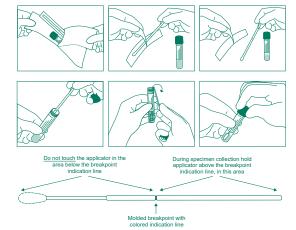
LIMITATIONS

- No Because calcium alginate swabs are toxic for many enveloped viruses⁵ and may interfere with immunofluorescence tests², they should not be used for specimen collection. Wooden shaft swabs may contain toxins and formaldehydes^{2,6} and should not be used. UTM kit is intended to be used with the medium tubes and swabs provided inside the kit. The use of tubes of medium or swabs from any other source may compromise product performance. 2 performance.

2

HPC170A Rev.01 Date 2019.09

Fig 1. Collection swab showing breakpoint indication line and area for holding the applicator



DISPOSAL

Waste must be disposed of in compliance with local legislation. Take the appropriate precautions for infected material if necessary.

QUALITY CONTROL

The UTM[®] System is tested to guarantee the absence of toxicity for the cellular lines used for the viral cultures and the ability to maintain the viability of viral, chlamydia and mycoplasma strains for up to 48 hours at 2-25°C in accordance with the methods described in CLSI M40-A2⁵.

RESULTS AND PERFORMANCE

The results obtained largely depend on proper and adequate specimen collection as well as the promptness with which the specimens are transported to the laboratory and analysed.

Viability studies were performed using UTM[®] with a panel of representative strains of the various families supported by the UTM[®]. The swabs that accompany each transport system were directly inoculated in triplicate with 100 µl of organism suspension. Subsequently, the swabs were inserted in the respective test tubes containing the transport medium and stored for 0 and 48 hours at 2-6°C and at controlled room temperature (20-25°C). At the time of processing, each swab was vortexed and at controlled room temperature (20-25°C). At the time of processing, each swab was vortexed for 20 seconds and removed from its transport medium test tube. At this point, an aliquot of the su-spension was inoculated into the cellular line (200 µl) or into the appropriate culture medium^{6,15}. All the cultures were processed using the standard laboratory culture technique^{6,15}. Organism viability was determined by fluorescent cell counting for viral and chlamydia strains and CFU counting for mycoplasma strains. The acceptability limits for time zero and for 48 hours were defined in accor-dance with the regulations M40-A2⁵. UTM[®] System preserved the viability of all the organisms tested for 48 hours at both controlled room temperature and in the refrigerator in the above described test conditions. The organisms evaluated and the results obtained are given in the table below

HPC170A Rev 01 Date 2019 09

Organism	ATCC number	Zero time	48 hours time 2-6 °C	48 hours time 20-24 °C
Herpes Simplex Virus Type 1	ATCC VR-539	++	+	+
Herpes Simplex Virus Type 2	ATCC VR-734	++	+	+
Respiratory Syncytial Virus	ATCC VR-1580	++	+	+
Coxsackie B1 Virus	ATCC VR-28	++	+	+
Chlamydia trachomatis	ATCC VR-880	++	+	+
Influenza A	ATCC-VR-1679	++	+	+
Cytomegalovirus	VR-977	++	+	+
Mycoplasma pneumonaie	ATCC 15331	++	+	+
Varicella-zoster virus	ATCC VR-1367	++	+	+
Chlamydia pneumoniae	ATCC VR-1360	++	+	+

+++=100% infected cells +++=75% infected cells ++= 50% infected cells +=25% infected cells NOTE: The HealthLink UTM[®] performance tests were conducted using laboratory strains and not human samples

TABLE OF SYMBOLS

See the table of symbols at the end of the instructions for use.

BIBLIOGRAPHY

- 1.
- RAPHY Gary W. Procop and Elmer W. Koneman, 2016. Color Atlas and Textbook of Diagno-stic Microbiology, Seventh edition. Wolters Kluwer Health. James H. Jorgensen, Michael A. Pfaller, Karen C. Carroll, Guido Funke, Marie Loui-se Landry, Sandra S. Richter, David W. Warnock, 2015. Manual of Clinical Microbio-logy, 11th Edition. ASM, Washington, DC. James Versalovic, Karen C. Carroll, Guido Funke, James H. Jorgensen, Marie Loui-se Landry, David W. Warnock, 2011. Cumitech 15A. Laboratory Diagnosis of Viral Infections. ASM, Washington, DC. Patricia Tille. 2014. Bailey & Scott's Diagnostic Microbiology, 13th Edition. Labora-tory Medicine. 2
- 3.
- 4.
- Particle Time 2014, Date y & Goott's Disgrissic Intercence, 1, 1997, 199 5.
- 6
- 7.
- 8. 9.
- Clinical and Laboratory Standards Institute (CLSI), 2006. M41-A Viral Culture; Approved guidelines. Wardford, A., M. Chernesky, and E. M. Peterson, 1999. Cumitech 19A, Laboratory Diagnosis of Chlamydia trachomatis Infections. ASM, Washington DC. 42CFR72. Code of Federal Regulations, Title 42, Volume 1, Part 72. Interstate Shipment of Etiologic Agents. J. Michael Miller, Shelley A. Miller, 2017. A Guide to Specimen Management in Clinical Microbiology, Third Edition. ASM, Washington DC. Centers for Disease Control and Prevention (CDC), 2016. Guide for Shipping In-fectious Substances 10 fectious Substances.
- Centers for Disease Control and Prevention (CDC), 2009. Biosafety in Microbiologi-cal and Biomedical Laboratories 5th Edition. World Health Organization 2015. Guidance on regulations for the Transport of In-11.
- 12.
- 13
- Voring Health Organization 2015. Collidance on regulations for the Transport of In-fectious Substances 2015 2016. Centers for Disease Control and Prevention (CDC), 2002. Screening Tests to De-tect Chlamydia trachomatis and Neisseria gonorrhoeae Infections 2002. Centers for Disease Control and Prevention (CDC), 2014. Recommendations for the Laboratory-Based Detection of Chlamydia trachomatis and Neisseria gonor-rhoeae 2014. 14. European Collection of Authenticated Cell Cultures (ECACC), Fundamental Tech-15.
- 16.
- European Collection of Authenticated Cell Cultures (ECACC). Fundamental Tech-niques in Cell Culture Laboratory Handbook, 3rd edition (Protocol 9 Detection of Mycoplasma by Culture Isolation). J.B. Mahony, M.A. Chernesky Effect of Swab Type and Storage Temperature on the Isolation of Chlamydia trachomatis from Clinical Specimens Journal of Clinical Microbiology, Nov. 1985, p. 865-867. S. Specter, R. L. Hodinka, S. A. Young. Clinical Virology Manual, fifth edition, 2016. Peter Daley, Santina Castriciano, Max Chernesky, Marek Smieja. Comparison of Flocked and Rayon Swabs for Collection of Respiratory Epithelial Cells from Unin-fected Volunteers and Symptomatic Patients. Journal of Clinical Microbiology, June 2006. 17. 18 2006.
- K. Loens, L. Van Heirstraeten, S. Malhotra-Kumar, H. Goossens, and M. leven. Optimal Sampling Sites and Methods for Detection of Pathogens Possibly Causing Community-Acquired Lower Respiratory Tract Infections. Journal of Clinical Micro-19
- biology, Jan. 2009. Richard Garceau MD, Danielle Leblanc RT, Louise Thibault MD, Gabriel Girouard MD, Manon Mallet PhD. Herpes simplex virus type 1 is the leading cause of genital herpes in New Brunswick. Can J Infect Dis Med Microbiol 2012. 20

5

HPC170A Rev.01 Date 2019.09

6

- Marek Smieja, Santina Castriciano, Susan Carruthers, Geoffrey So, Sylvia Chong, Kathy Luinstra, James B. Mahony, Astrid Petrich, Max Chernesky, Mario Savarese, and Daniele Triva. Development and Evaluation of a Flocked Nasal Midturbinate Swab for Self-Collection in Respiratory Virus Infection Diagnostic Testing. Journal of Clinical Microbiology, Sept. 2010. Donghyok Kwon, Kyeongcheol Shin, Mihwa Kwon, Hee-Bok Oh, Chun Kang, Joo-Yeon Lee. Development and Evaluation of Rapid Influenza Diagnostic Test for the Pandemic (H1N1) 2009 Virus. JCM Accepts, published online ahead of print on 27 October 2010. Bhupesh K. Prusty, Christine Siegl, Petra Hauck, Johannes Hain, Suvi J. Korhonen, Eija Hiltunen- Back, Mirja Puolakkainen, Thomas Rudel. Chlamydia trachomatis Infection Induces Replication of Latent HHV-6. Plos One, April 2013. Ellen Vancutsem, Oriane Soetens, Maria Breugelmans, Walter Foulon, Anne Naessens Modified Real-Time PCR for Detecting, Differentiating, and Quantifying Ureaplasma urealyticum and Ureaplasma parvum. The Journal of Molecular Diagnostics, Vol. 13, No. 2, March 2011. 21
- 22 23
- 24
- 2. March 2011.
- 2, march 2017. Turkiya Al-Siyabi, Khalifa Binkhamis, Melanie Wilcox, Sallene Wong, Kanti Pabbaraju, Raymond Tellier, Todd F. Hatchette, and Jason J. Le Blanc. A cost-effective real-time PCR for the detection of adenovirus from viral swabs. Al-Siyabi et al. Virology Journal 25 2013

HPC170A Rev.01 Date 2019.09

Symbol	Meaning		
	Manufacturer		
IVD	In vitro diagnostic device		
C € 0123	Identification number of notified body		
STERILE EO	Sterilized using ethylene oxide		
STERILE R	Sterilized using irradiation		
2	Do not reuse		
REF	Catalogue number		
	Temperature limitation		
	Use by de		
Ĩ	Consult Instructions for Use		
A	Peel		
LOT	Batch code (Lot)		
Σ	Contains sufficient for <n> tests</n>		
	Do not use if package is damaged		
Rx Only	Prescription Device Labeling Statement		
	Country of manufacture		

Copan Italia S.p.A. Via F. Perotti, 10 25125 Brescia, Italy

8

Distributed by: Hardy Diagnostics 1430 W McCoy Ln, Santa Maria, CA 93455, USA Tel: 1 800-266-2222 - www.hardydiagnostics.co

